ABERRANT EXPRESSION OF GLUCAGON RECEPTORS IN ADRENAL GLANDS OF A PATIENT WITH CUSHING’S SYNDROME AND ACTH-INDEPENDENT MACRONODULAR ADRENAL HYPERPLASIA

VALERIA DE MIGUEL¹, MARIA ANA REDAL², MARIA LORENA VIALE³, MARIANO KAHAN⁴, MARIELA GLEREAN⁵, AXEL BESKOW⁶, PATRICIA FAINSTEIN DAY⁷

¹Servicio de Endocrinología, Metabolismo y Medicina Nuclear, ²Unidad de Medicina Molecular y Genómica, ³Departamento de Cirugía, Hospital Italiano de Buenos Aires

Abstract  Adrenocorticotropin (ACTH) independent bilateral macronodular adrenal hyperplasia (AIMAH) is a rare cause of Cushing’s syndrome, characterized by bilateral adrenal lesions and excess cortisol production despite ACTH suppression. Cortisol synthesis is produced in response to abnormal activation of G-protein-coupled receptors, such as gastric inhibitory peptide, vasopressin, beta adrenergic agonists, LH/hCG and serotonin receptors. The aim of this study was to analyze the expression of glucagon receptors in adrenal glands from an AIMAH patient. A patient with ACTH-independent Cushing’s syndrome and bilateral macronodular adrenal hyperplasia was screened for altered activation of adrenal receptors by physiological (mixed meal) and pharmacological (gonadotrophin releasing hormone, ACTH and glucagon) tests. The results showed abnormally high levels of serum cortisol after stimulation with glucagon. Hypercortisolism was successfully managed with ketoconazole treatment. Interestingly, a 4-month treatment with a somatostatin analogue (octreotide) was also able to reduce cortisol secretion. Finally, Cushing’s syndrome was cured after bilateral adrenalectomy. Abnormal mRNA expression for glucagon receptor in the patient’s adrenal glands was observed by Real-Time PCR procedure. These results strongly suggest that the mechanism of AIMAH causing Cushing’s syndrome in this case involves the illicit activation of adrenal glucagon receptors. This is the first case reported of AIMAH associated with ectopic glucagon receptors.

Key words: Cushing’s syndrome, adrenocorticotropin, ACTH-independent bilateral macronodular adrenal hyperplasia (AIMAH), glucagon receptor, illicit receptor

Recibido: 19-I-2010 Aceptado: 30-III-2010

Postal address: Dra. Patricia Fainstein Day, Servicio de Endocrinología, Metabolismo y Medicina Nuclear, Hospital Italiano, Av. Gascón 450, 1181 Buenos Aires, Argentina
Fax: (5411) 4959-0323 e-mail: patricia.fainstein@hospitalitaliano.org.ar
Cushing’s syndrome (CS) results from chronic exposure to high serum cortisol levels and other adrenal steroids. Bilateral adrenal lesions occur in 10-15% of adrenal CS, including corticotropin (ACTH)-independent macronodular adrenal hyperplasia (AIMAH). The excess cortisol secretion, which leads to progressive inhibition of ACTH release is under control of ligands that bind to illegitimate G-protein-coupled receptors (GPCRs) in the adrenal glands (AG). The expression of ectopic receptors like gastric inhibitory polypeptide (GIP), beta adrenergic, vasopressin V2-V3, serotonin (5-HT) receptors, and abnormally active eutopic receptors like vasopressin V1, luteinizing hormone/human gonadotrophin (LH/hCGR) and serotonin (5-HT) receptors have being described.

We present the study of a patient evaluated for Cushing’s syndrome. A 64-year old man reported a history of obesity, severe arterial hypertension, diabetes mellitus, congestive heart failure, mild chronic renal insufficiency, weakness and depression. Physical examination showed clinical signs of Cushing’s syndrome including central abdominal obesity, thin skin, dorsocervical fat pad, red dish purple striae, proximal muscle weakness, and ecchymoses. Biochemical studies revealed: 24-hour urinary free cortisol (UFC) 250 µg (normal up to 100), late-night free salivary cortisol: 24 nmol/l (0.7-5) and serum cortisol after 1 mg overnight dexamethasone suppression test: 32 µg/dl (up to 5). Morning plasma ACTH was undetectable: less than 10 pg/ml (10-46), confirmed in three assays. An abdominal computed tomography (CT) scan showed bilateral macronodular adrenal hyperplasia. The right gland measured 11 x 4 cm, and the left, 8 x 5 cm. To screen for aberrant adrenal expression of GPCRs we performed some of the stimulation tests, according to the protocol suggested by Lacroix et al. The strategy is based on stimulation tests with different hormones (endogenous or exogenous) for potential aberrant receptors, while monitoring serum levels of cortisol. First, a mixed meal was given to assess the presence of GIP, also 250 µg of ACTH was administrated intravenously. During the second day, 100 µg of gonadotrophin-releasing hormone (GnRH) was administrated intravenously and on the last day, 1 mg of glucagon was administrated intramuscularly. Serial measurement of cortisol levels were performed every 30 minutes intervals during 3 hours following the intervention. ACTH was not measured during the tests. The results showed an increased cortisol response to stimulation with glucagon (Table 1). Cortisol response to ACTH is always present in AIMAH patients. After 12-months of successful treatment with ketoconazole, the patient was submitted to bilateral adrenal surgery. Right adrenal excision had to be postponed because of harmful bleeding. Based on biochemical studies that showed cortisol response after exogenous glucagon administration, we decided to treat the patient with octreotide (20 mg/month) in order to control cortisol secretion as well as to slow down adrenal hyperplasia. After four months of octreotide treatment cortisol secretion could be partially controlled (UFC: 140-200 µg/24 h) and the surgical excision of the right adrenal could be easily made without abnormal bleeding. Interestingly, exogenous glucagon stimulus during octreotide treatment did not elicit significant cortisol response. We have no argument to explain this finding.

Pathological studies revealed adrenal nodular hyperplasia. The patient is under hydrocortisone (30 mg) and fludrocortisone (0.015 mg) treatment.

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Table 1: Stimuli tests for cortisol

<table>
<thead>
<tr>
<th>Stimuli tests</th>
<th>Basal</th>
<th>30 min</th>
<th>60 min</th>
<th>90 min</th>
<th>120 min</th>
<th>180 min</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mixed meal</td>
<td>35</td>
<td>28.7</td>
<td>30.6</td>
<td>30.3</td>
<td>31</td>
<td></td>
</tr>
<tr>
<td>LHRH, 100 µg iv</td>
<td>32</td>
<td>30.6</td>
<td>31.2</td>
<td>31.6</td>
<td>31</td>
<td></td>
</tr>
<tr>
<td>ACTH, 250 µg iv</td>
<td>29</td>
<td>63.7</td>
<td></td>
<td>77</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Glucagon, 1 mg im</td>
<td>31.5</td>
<td>25.6</td>
<td>27.6</td>
<td>86.7</td>
<td>94.8</td>
<td>112</td>
</tr>
</tbody>
</table>

Fig. 1. Electrophoresis in agarose gel stained by ethydium bromide. Control beta actin, a PCR product of 122 bp; line 1: patient (C2); line 2: hepatocarcinoma cell line (CL); line 3: normal adrenal gland (C1); line 4: negative control assay (B).
We studied adrenal glucagon receptor (GCGR) mRNA expression in normal adrenal glands and in the patient adrenal gland by Real Time PCR (RT-PCR) assays. In order to do this, total RNA was extracted from the patient right adrenal gland and purified with Trizol RNA purification kit (Invitrogen, USA). The corresponding cDNA was obtained using RT polymerase Improm II (Promega) and used as template for RT-PCR assay. cDNA obtained from an hepatocarcinoma cell line was used as a positive control for GCGR expression, and cDNA from normal adrenal gland as a negative control. Beta-actin was used as an amplifications control. RT-PCR was performed in a Light-Cycler 2.0 System (Roche Applied Science, USA) with the specific GCGR primers forward (5’-GCCTGACCCTCATCCCTCG-3’) and reverse (5’-TAGAGGACAGCCACCAGCAG-3’) using the Taq Platinum polymerase (Invitrogen) and SYBR Green I (Invitrogen) to follow the kinetic reaction. Cycling conditions were: one cycle at 94 °C for 10 min and 45 cycles at 94 °C for 10 s, 57 °C for 10 s and 72 °C for 10 s. As shown in Fig. 1, the 148 bp PCR product corresponding to GCGR mRNA was present in the patient’s adrenal gland but absent in the normal control gland.

In conclusion, we suggest that cortisol hypersecretion was related to ectopic expression of glucagon receptor in the adrenal gland of this patient. The cortisol response to exogenous glucagon stimulation and the finding of glucagon receptor mRNA strongly support this hypothesis. This is the first case reported of AIMAH associated with ectopic glucagon receptors.

Conflict of interest: None to declare.

References